

Isoprenoids

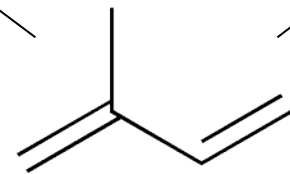
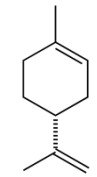
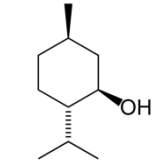
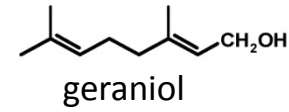
monoterpenoids

polyterpenoids

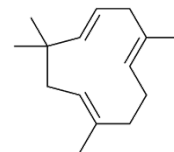
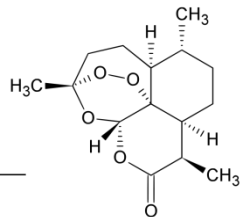
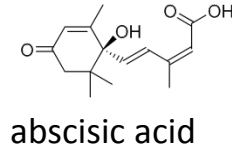


rubber

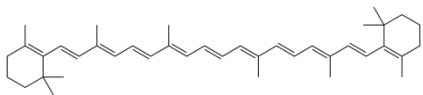
hemiterpenoids



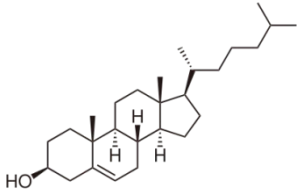
sesquiterpenoids



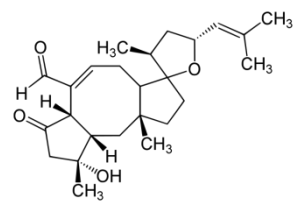
tetraterpenoids



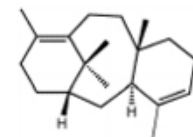
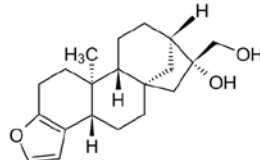
triterpenoids



sesterterpenoids



diterpenoids



n > 8

n = 1

n = 2

n = 8

n = 3

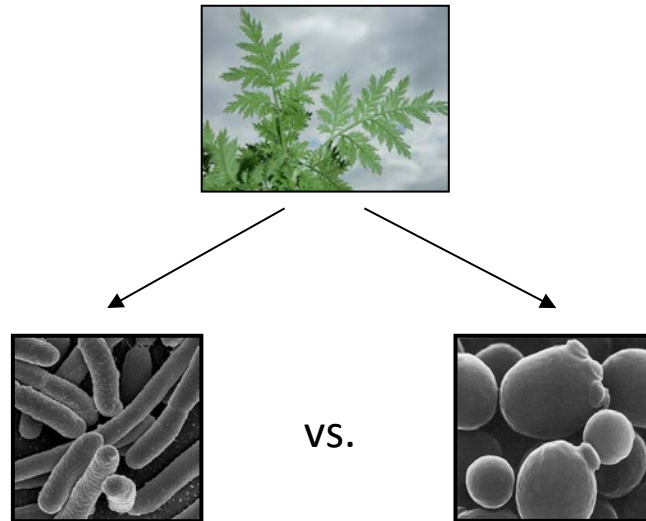
n = 6

n = 5

n = 4

Isoprenoids

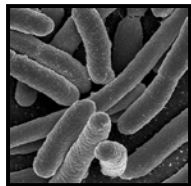
Pathway transfer from plant to microorganism



Isoprenoids

2 metabolic pathways

non-mevalonate pathway
(DXP pathway, MEP pathway)

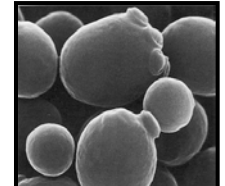


eubacteria
plants (plastides)
apicomplexa

precursors:
pyruvate + glyceraldehyde-3-phosphate

1 glucose per IPP

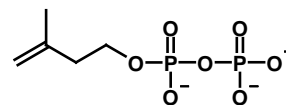
mevalonate pathway
(MVA pathway)



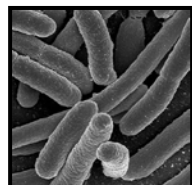
eukaryotes
plants (cytosol)
archaea, few eubacteria

precursor:
acetyl-CoA

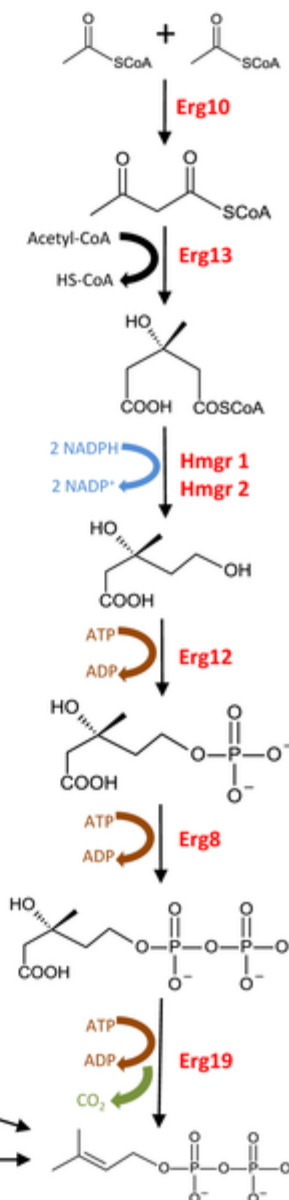
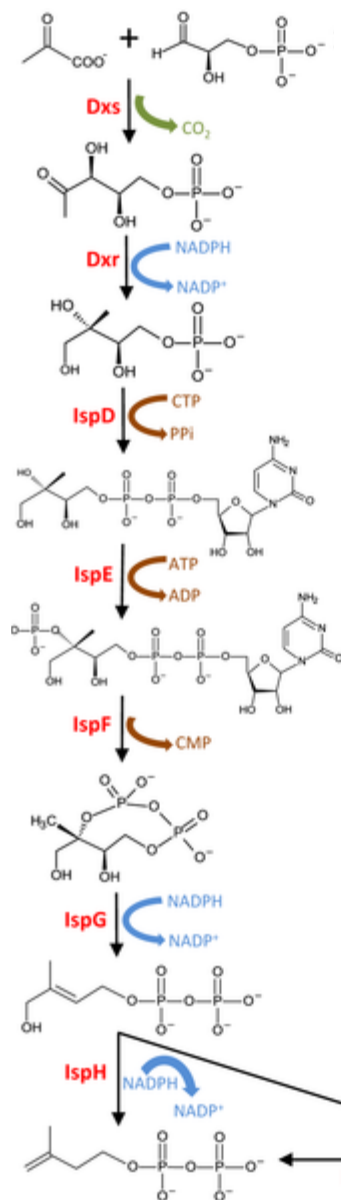
1.5 glucose per IPP



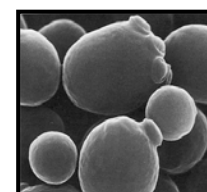
IPP



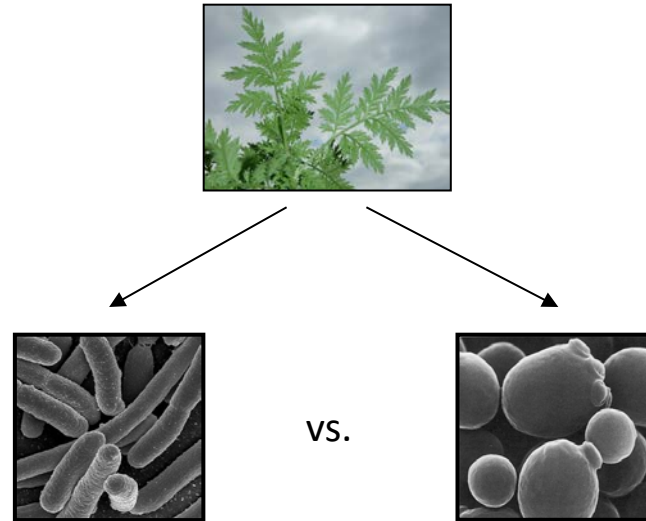
MEP pathway



MVA pathway



Isoprenoids



Advantage *E. coli*: more efficient pathway -> higher theoretical yields

Advantage *S. cerevisiae*: better expression host for auxiliary enzymes
(P450 monooxygenases)

=> choice may depend on product

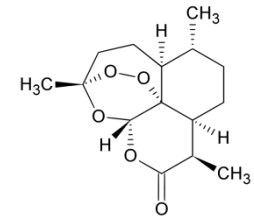
Artemisinin

Antimalarial drug precursor

Derivatives used in antimalarial combination therapies

Derived from sweet wormwood plant (*Artemisia annua*)

Fluctuating price and availability



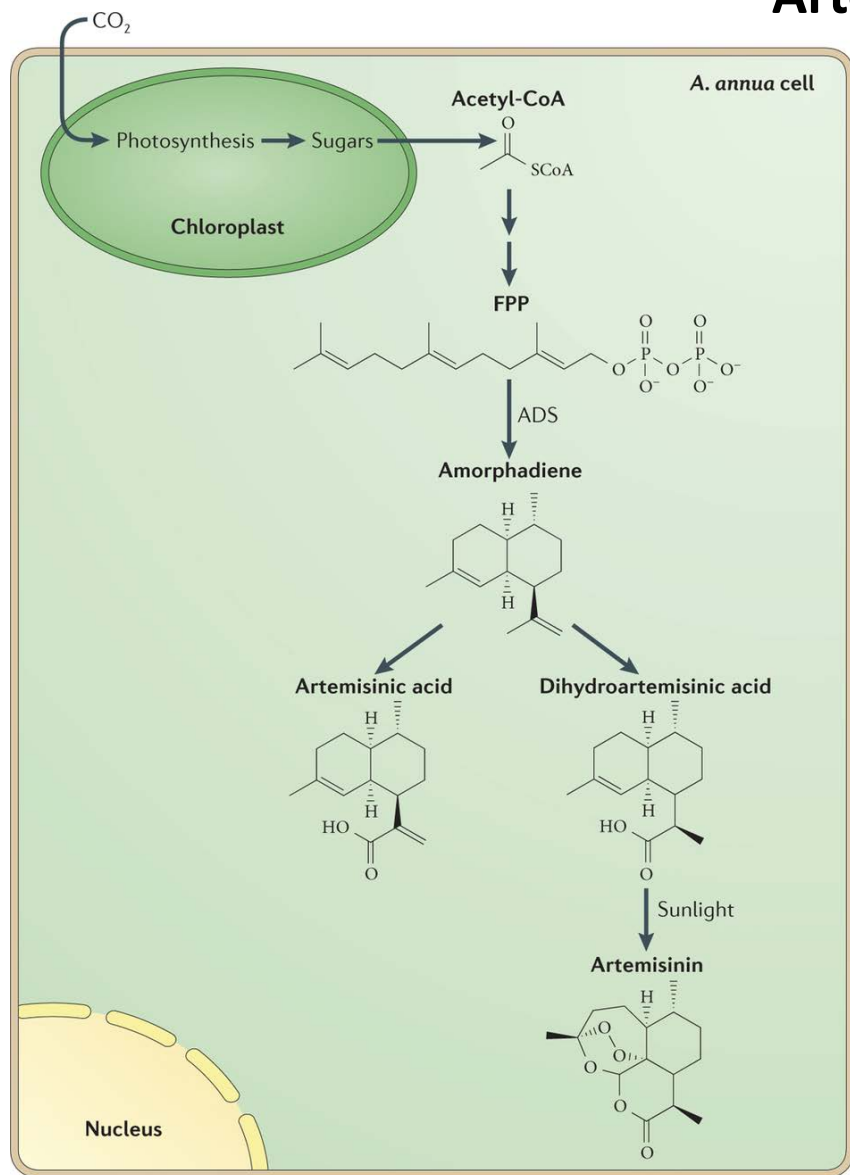
⇒ Semi-synthetic artemisinin project:

Started in 2004

Funded by Bill and Melinda Gates Foundation

Collaboration between UC Berkeley, Amyris, Institute for OneWorld Health

Artemisinin



Plant pathway:

Conversion of FPP to amorphadiene via amorphadiene synthase (ADS)

2 oxidation products: artemisinic acid, dihydroartemisinic acid

spontaneous conversion of dehydroartemisinic acid to artemisinin

Project aim:

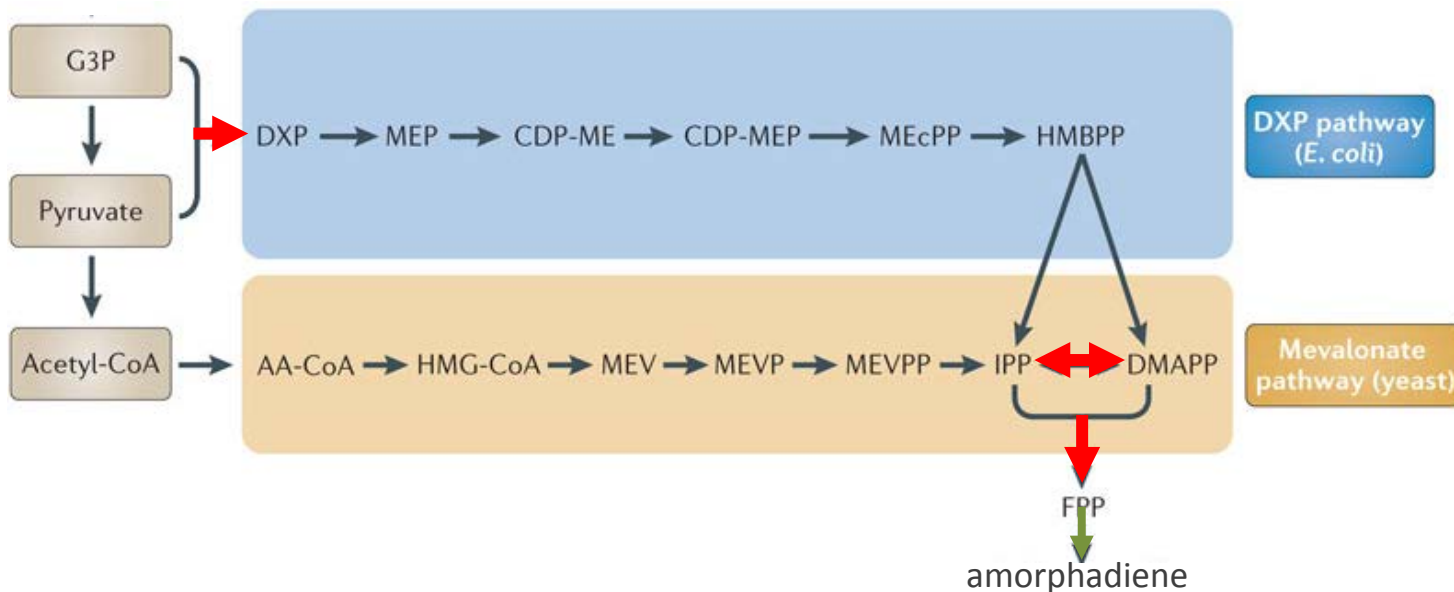
Production of artemisinic acid in *E. coli*

Chemical conversion to artemisinin

Artemisinin

1. Amorphadiene production in *E. coli*

- Use of plant terpene synthases with limited success
- Overexpression of rate-limiting DXP pathway enzymes with limited success (probably due to internal regulatory mechanisms)



Artemisinin

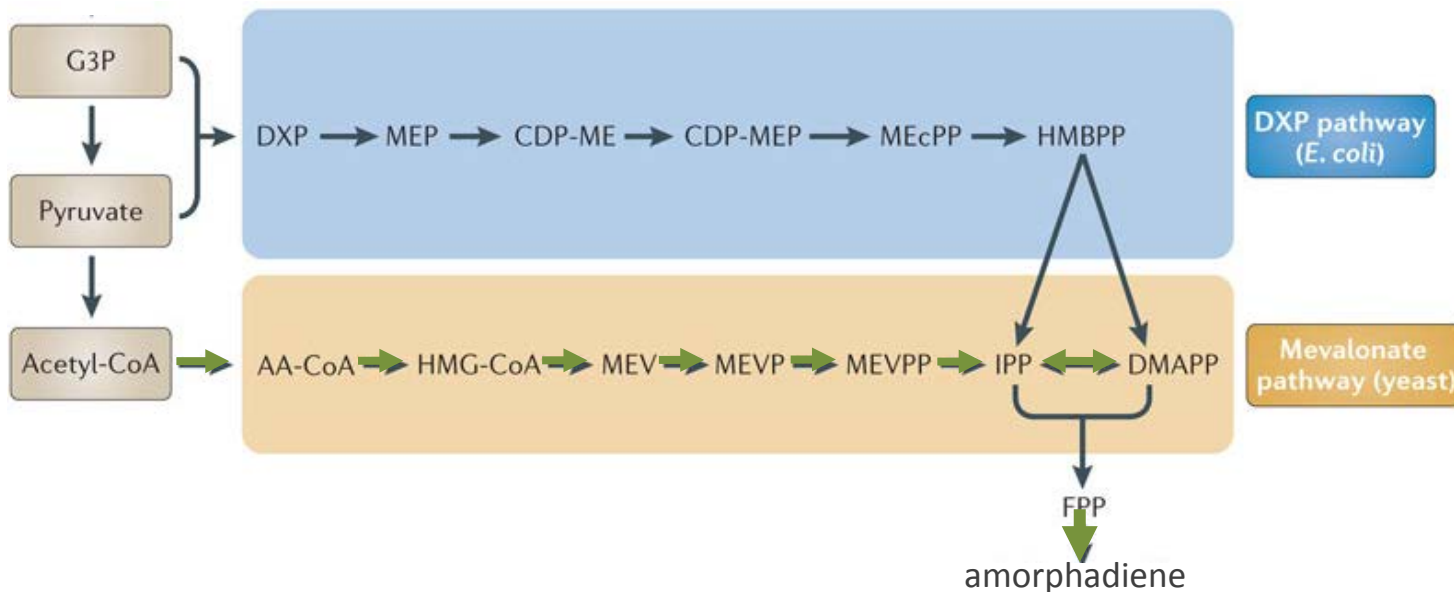
1. Amorphadiene production in *E. coli*

- Use of plant terpene synthases with limited success
- Overexpression of rate-limiting DXP pathway enzymes with limited success

⇒ Expression of codon-optimised ADS

⇒ Expression of yeast MVA pathway

-> 0.5 g/l amorphadiene



Artemisinin

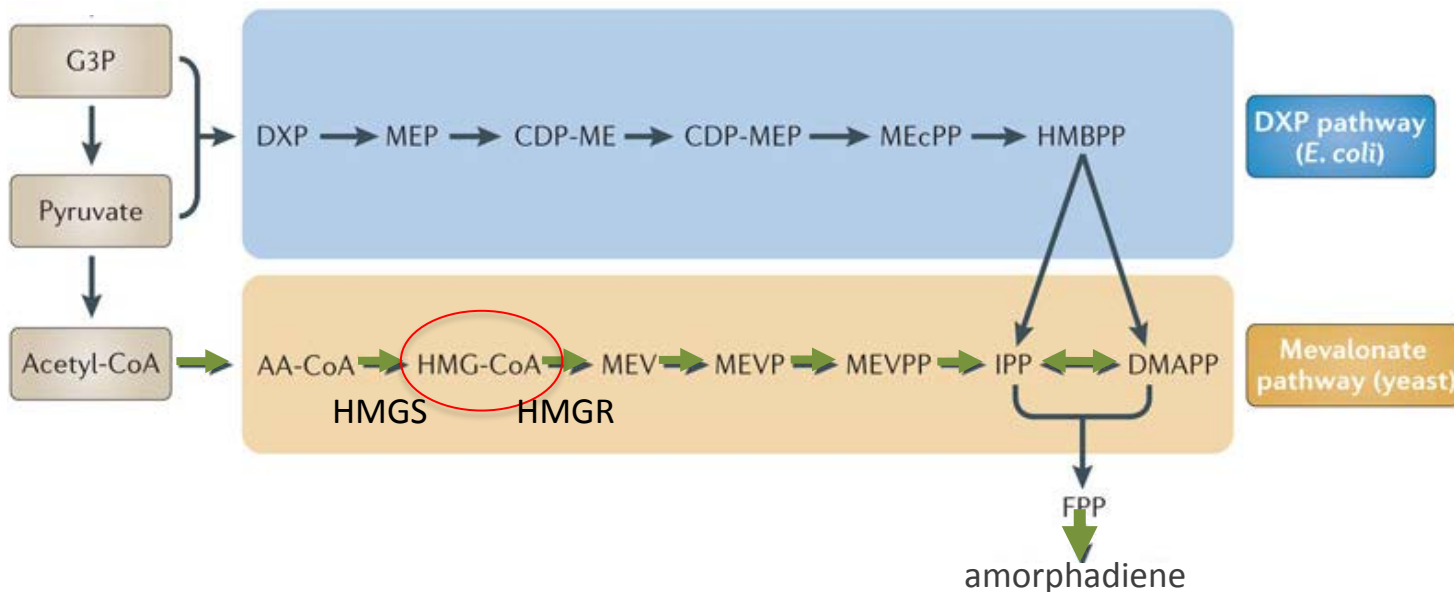
1. Amorphadiene production in *E. coli*

Problem: Imbalanced enzyme expression leading to HMG-CoA accumulation and growth inhibition

Solution: Balanced expression of HMGS and HMGR, replacement of yeast enzymes with bacterial enzymes

Fermentation optimisation

-> 25 g/l amorphadiene



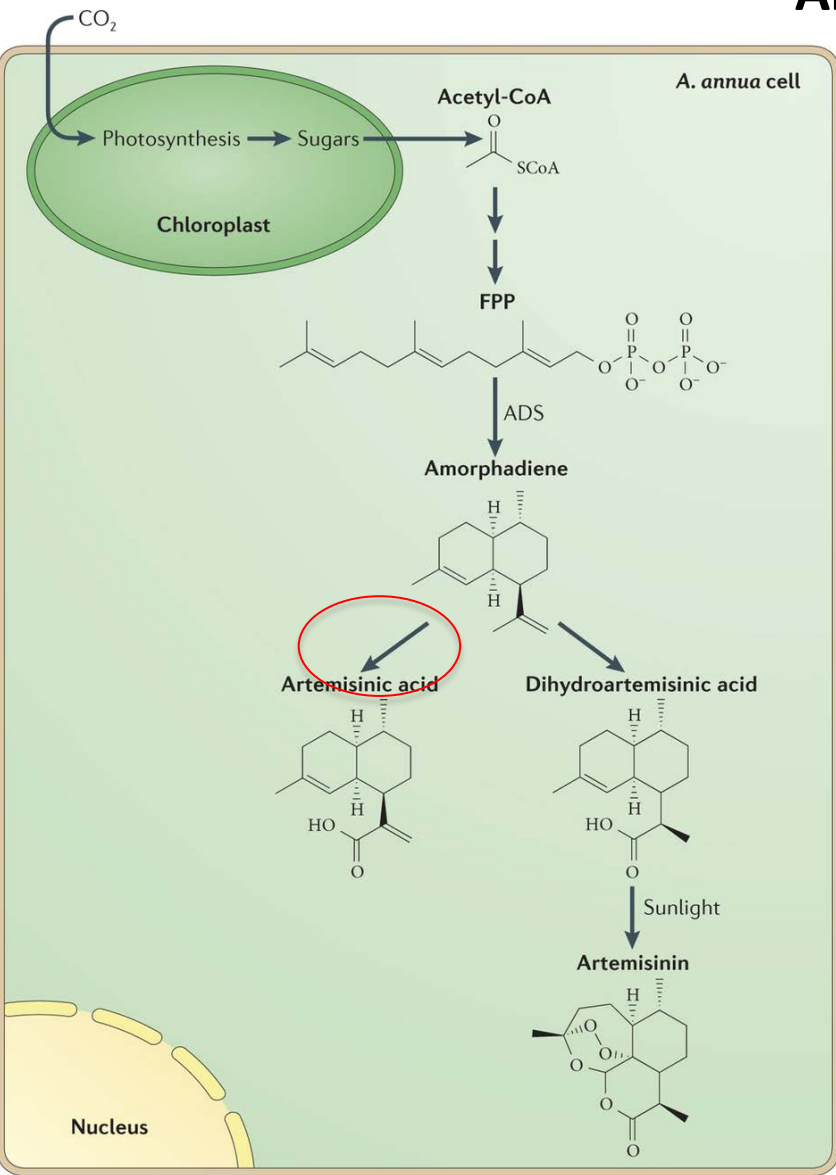
Artemisinin

2. Switch to yeast

a) Identification of enzyme converting amorphaadiene to artemisinic acid:

CYP71AV (P450 monooxygenase)

-> 100 mg/l artemisinic acid in primarily engineered strain

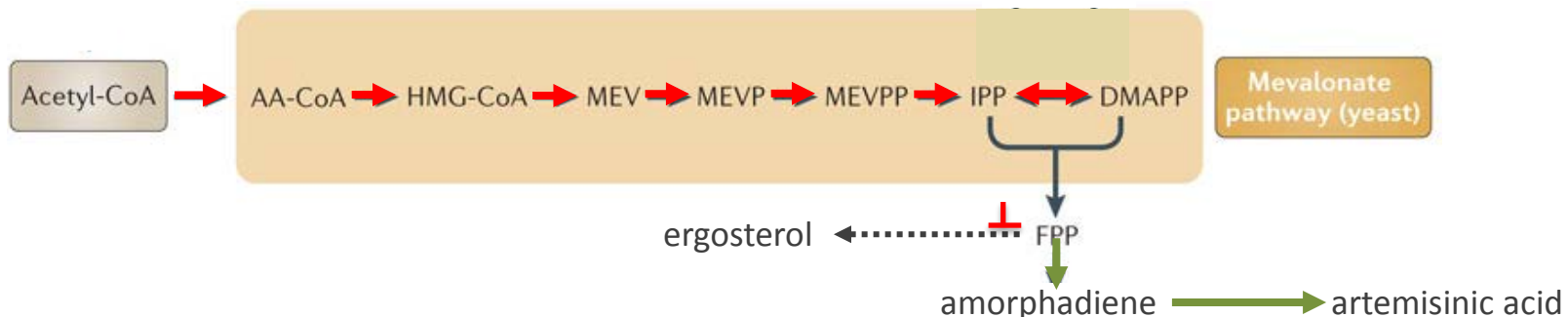


2. Switch to yeast

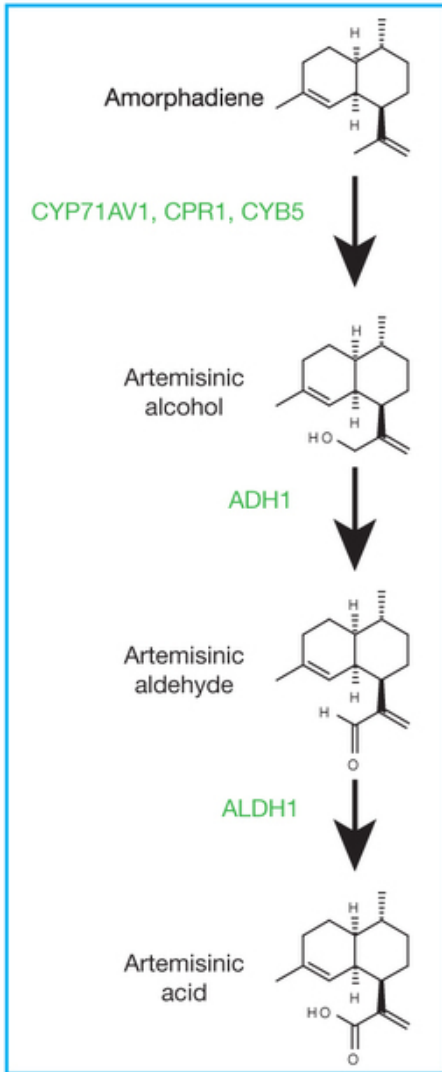
b) Pathway optimisation

- Overexpression of all MVA pathway genes
- Downregulation of first step of ergosterol formation (gene under regulation of methionine- or copper-regulatable promoter)
- Fermentation optimisation → 40 g/l amorphadiene

but: amorphadiene production 10x higher than artemisinic acid production



Artemisinin



2. Switch to yeast

c) Optimisation of oxidation step

Engineered strains with low viability due to oxidative stress
(poor coupling between reductase CPR1 and CYP71AV1)

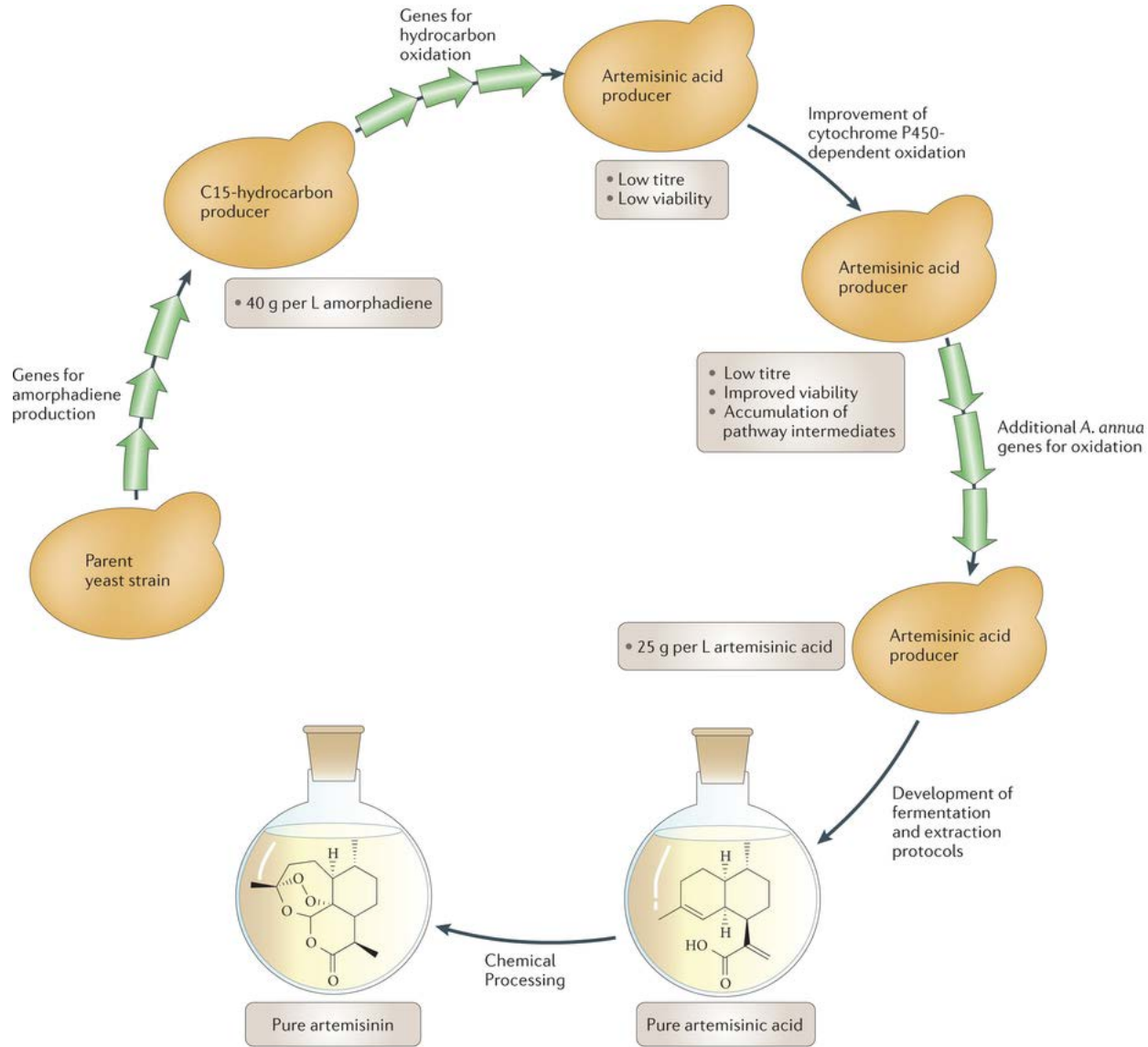
⇒ Reduction of CPR1 expression, additional expression of cytochrome b₅

Accumulation of oxidation intermediates

⇒ Additional expression of *A. annua* aldehyde dehydrogenase ALDH1 and alcohol dehydrogenase ADH1

-> 25 g/l artemisinic acid

Artemisinin



Conclusion (Part I)

Yeast as a suitable host for production of high-value compounds

Engineering strategies often product specific and time consuming

⇒ Need for better predictability, universal strategies, platform strains

Systems biology for the improvement of microbial fermentations

Verena Siewers

Systems and Synthetic Biology
Chalmers University of Technology

Campinas, 12 October 2014

The metabolic engineering cycle

